

Atty. Dkt. No. 071957-0903

**Amendments to the Specification:**

Please amend the specification as follows:

Please replace the paragraphs starting at page 20, line 6, (the 2<sup>nd</sup>, 3<sup>rd</sup>, 4<sup>th</sup>, 5<sup>th</sup>, 6<sup>th</sup>, 7<sup>th</sup>, and 8<sup>th</sup> paragraphs), with the following rewritten paragraphs:

Figure 1 shows a comparison of flow cytometric detection of interleukin-2 expression in human peripheral blood mononuclear cells stimulated by exposure to phorbol myristic acetate and ionomycin, using an amplification medium comprising 20 mM glycylglycine, pH 8.0, 1 M NaCl, 0.01% peroxide, and 100 µg/mL fluorescein-tyramide. Filled histograms represent cells stained with control Ig, and open histograms represent cells stained with monoclonal anti-human interleukin 2 antibodies.

Figure 2 shows a comparison of flow cytometric detection of bcl-2 expression in CEM cells, using an amplification medium comprising 50 mM (N-tris (hydroxymethyl) methyl-3-aminopropanesulfonic acid ("TAPS"), pH 8.0, 1.5 M NaCl, 0.01% peroxide, and 100 µg/mL fluorescein-tyramide. Filled histograms represent cells stained with control Ig, and open histograms represent cell stained with anti-bcl-2.

Figure 3 shows a comparison of flow cytometric detection of bcl-2 expression in K562 cells, using an amplification medium comprising 50 mM (N-tris (hydroxymethyl) methyl-3-aminopropanesulfonic acid ("TAPS"), pH 8.0, 1.5 M NaCl, 0.01% peroxide, and 100 µg/mL fluorescein-tyramide. Filled histograms represent cells stained with control Ig, and open histograms represent cells stained with anti-bcl-2.

Figure 4 shows a comparison of flow cytometric detection of bcl-2 expression in HUT-102 cells, using an amplification medium comprising 20 mM glycylglycine, pH 8.0, 1 M NaCl, 0.01% peroxide, and 100 µg/mL fluorescein-tyramide. Filled histograms represent cells stained with control Ig, and open histograms represent cells stained with anti-bcl-2.

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Figure 5 shows a comparison of flow cytometric detection of bcl-2 expression in JY(LCL) cells, using an amplification medium comprising 20 mM glycylglycine, pH 8.0, 1 M NaCl, 0.01% peroxide, and 100 µg/mL fluorescein-tyramide. Filled histograms represent cells stained with control Ig, and open histograms represent cell stained with anti-bcl-2.

Figure 6 shows a comparison of flow cytometric detection of Epstein-Barr Virus LMP-1 expression in JY-LCL cells, using an amplification medium comprising 50 mM (N-tris (hydroxymethyl) methyl-3-aminopropanesulfonic acid ("TAPS"), pH 8.0, 1.5 M NaCl, 0.01% peroxide, and 100 µg/mL fluorescein-tyramide. Filled histograms represent cells stained with control Ig, and open histograms represent cells stained with anti-bcl-2.

Figure 7 shows a comparison of flow cytometric detection of bcl-2 expression in CEM cells, using a primary FITC-conjugated anti-bcl-2 antibody. The term "standard staining" refers to direct detection of the primary antibody, while the term "enzymatic amplification" refers to the deposition of tyramide catalyzed according to the methods described herein, using a secondary horseradish peroxidase-conjugated anti-FITC secondary antibody, followed by an amplification medium comprising 20 mM glycylglycine, pH 8.0, 1 M NaCl, 0.01% peroxide, and 100 µg/mL fluorescein-tyramide. Filled histograms represent cells stained with control Ig, and open histograms represent cells stained with anti-bcl-2.

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**Amendments to the Drawings:**

The drawing sheets attached in connection with the above-identified application containing Figure(s) 1-7 are being presented as new formal drawing sheets to be substituted for the previously submitted drawing sheets. Figures 1-7 have been amended to delete excess text. Appended to this amendment is an annotated copy of the previous drawing sheets which have been marked to show changes presented in the replacement sheets of the drawing.